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PRIMER NOTE

DEVELOPMENT AND CHARACTERIZATION OF MICROSATELLITE LOCI FOR *LINDERA GLAUCA* (LAURACEAE)¹

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- Premise of the study: Microsatellite primers were developed to investigate population genetic structure in Lindera glauca (Lauraceae).
- Methods and Results: Twenty-five microsatellite primers were developed and optimized for L. glauca using Illumina's Solexa sequencing technology. These novel primers were found to be polymorphic in nine wild L. glauca populations with 81 total alleles confirmed and genotyped via capillary gel electrophoresis. The total number of alleles, observed heterozygosity, and expected heterozygosity for each population ranged from one to four, from 0.00 to 0.90, and from 0.00 to 0.79, respectively. In addition, the 25 primers were tested in 10 additional individuals of the related species L. communis, and all but four markers showed good amplification results.
- Conclusions: This set of microsatellite primers is the first specifically developed for L. glauca and will facilitate studies of genetic diversity and evolution among populations of this species.

Key words: genetic diversity; Lauraceae; Lindera glauca; microsatellite; polymorphism.

Lindera glauca Blume is a deciduous shrub or small tree that belongs to the family Lauraceae. It is extensively distributed in mountainous regions at low altitudes in central and southern China and is also found in Japan, Korea, and Taiwan. It is of potentially great economic value and ecological importance owing to its various valuable properties, including its natural abundance, the medicinal value of its leaves and roots, its high-quality wood, and the wide applications of its volatile oil in the biochemical and medicinal industries (Liu et al., 1992; Seki et al., 1994; Wang et al., 1994, 2011; Sun et al., 2011; Huh et al., 2014). However, few studies have investigated its population genetic diversity and genetic relationships among germplasms and breeding populations. Male individuals of L. glauca trees are very rare in China, and only female individuals are found in Japan (Dupont, 2002), although male individuals have been known from continental Asia in the past several decades (Wang, 1972; Li, 1982). Consequently, understanding the genetic diversity of this species is relevant to the utilization and conservation of its germplasm resources, to population genetic studies, and to the evolution of apomixis in this dioecious species.

Microsatellites, or simple sequence repeats (SSRs), have been widely used as genetic markers owing to their multiallelic nature, codominant inheritance, and thorough genome coverage (Powell et al., 1996). They are a powerful tool and an effective

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way to analyze population genetic structure, marker-assisted breeding, gene flow, levels of inbreeding, and germplasm identification (Varshney et al., 2005). However, no studies have previously published SSR markers for this species. Therefore, we used a next-generation transcriptome sequencing approach (Illumina's Solexa sequencing technology) to develop microsatellites specifically for *L. glauca*.

METHODS AND RESULTS

Plant materials and DNA extraction—Leaves and fruits of wild *L. glauca* were collected from nine locations in China in 2014 and 2015 (Appendix 1). Genomic DNA was extracted from the leaves of one individual from each of nine total populations using a modified cetyltrimethylammonium bromide (CTAB) method (Doyle and Doyle, 1987).

Development of SSRs and primer design—In this study, we used transcriptome data from Niu et al. (2015) to develop microsatellite markers. We used the 163,427 unigenes from the transcriptome data for SSR exploitation using QDD version 3.1 software (Meglécz et al., 2014) with at least five, five, four, four, three, and two SSR motif repeat units for di-, tri-, tetra-, penta-, hexa-, and hep-tanucleotide and higher-order nucleotides, respectively. A total of 8969 putative SSRs (excluding mononucleotide repeats) were detected, with the majority of repeats being dinucleotide (66.83%), followed by trinucleotide (33.77%), tetra-nucleotide (1.87%), pentanucleotide (0.50%), and hexanucleotide (1.04%). With this detailed information, the program PRIMER 5 (PRIMER-E, Auckland, New Zealand) was then used to design 27,350 primer pairs with primer lengths of 18–25 bp, amplification product sizes of 100–400 bp, GC contents from 40% to 60%, and annealing temperatures ranging from 55°C to 65°C.

PCR amplification and fragment analysis—An initial polymorphism screening of 120 primer pairs, including 50 primer pairs for dinucleotide motifs, 40 for trinucleotide motifs, 15 for tetranucleotide motifs, 10 for pentanucleotide motifs, and five for hexanucleotide motifs, was performed using

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TABLE 1. C	Characteristics of 25	5 microsatellite	loci developed	for Lindera glauca.
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Locus		Primer sequences $(5'-3')$	Fluorescent dye	Repeat motif	Allele size (bp)	$T_{\rm a}(^{\circ}{\rm C})$	GenBank accession no.
XBLG-013	F:	CGAGGGAGAGATCGACGC	FAM	(AG) ₅	190	58	KX545436
	R:	ATGGCACCACGAAGTGTGTT					
XBLG-033	F:	CGGGATGACAATTTGCATGT	HEX	(AG) ₅	259	58	KX545437
	R:	TGGAGCAGATTATGGTTTCCA					
XBLG-036	F:	CATCACCTCCCTCAAATCCC	FAM	$(AG)_7$	263	58	KX545438
	R:	GTTTCCGAAATTCTCGAGGC			101	50	
XBLG-049	F:	TTTCACAACCAGGGTGGCTA	TAM	$(AC)_6$	191	58	KX545439
VDL C 051	R:	CACTGGGACTAAGACACGGC	11FW		140	55	1232545440
XBLG-051	F.:	CAAACAGAACCAAGACA'I'CCAA	HEX	$(AIAC)_6$	148	55	KX545440
VDI C 052	K:	ATGGAGGAGCATGATTCGAG	TAM	(AT)	260	55	VV545441
ABLG-055	Ľ:		IAM	$(AI)_7$	208	55	KA343441
VPI C 055	K: E.		EAM	$(\Lambda \Lambda G)$	226	55	VV545442
ABLU-033	г: р.		PAM	$(AAO)_5$	230	55	KAJ4J442
XBI G-056	г. г.		ROX	(\mathbf{AG})	283	55	KX545443
ADLO-050	г. В.		КОХ	(AO)8	205	55	117734345
XBLG-058	F:	AGTCCAGGCTAACCAGACTCC	HEX	(AAC)	277	55	KX545444
1111110 000	R:	CCCAGTTTGCCAGGTAAGAA	11211	(1110)0	_,,	00	
XBLG-060	F:	ATTCCACCCATTCCCTTCTT	FAM	(AAG) ₆	197	55	KX545445
	R:	GATTCTAAGAAGAAGAAGAAGTACCC		(-)0			
XBLG-062	F:	AACATCATTCCCTCCATCCA	ROX	(AATCC) ₅	192	55	KX545446
	R:	CCAGCCAGTTAGGGTTTCAC					
XBLG-063	F:	CATGGCAACGCAAATCCTAT	TAM	(ATC) ₆	196	55	KX545447
	R:	CTAGATCCTTTGGCCATGTTT					
XBLG-066	F:	GTCGACGAGGACGAGGAC	HEX	(CCG) ₅	187	55	KX545448
	R:	TCGAATGAGGAAAGTTTGGC					
XBLG-073	F:	ACCACAAAGATAAGCTACAATGC	FAM	$(ACGC)_5$	219	55	KX545449
	R:	GGGCCTTAATGTCTATGGCA					
XBLG-076	F:	GGATGCTCTAAGGTGCTTGC	ROX	$(AG)_7$	182	55	KX545450
	R:	GGAATCGCCATTCTCCCT		(199	1.60		
XBLG-082	F:	TGTGGAAACAGAACCCATGA	TAM	$(AGC)_5$	168	55	KX545451
VDI C 092	R:		LIEV		107	55	WW545450
ABLG-085	Ľ:		HEA	$(AAG)_5$	180	55	KA343432
VDI C 084	K: E.		EAM	(ΛGG)	144	55	VV545452
ABLU-064	г: р.		PAM	$(AOO)_5$	144	55	KAJ4J4J3
XBI G-086	F.	TTCCCACTACCCTTTCATCC	TAM	(ACC)	190	55	KX545454
ADEG 000	E.	CCCATCATCAATGTGGTTATAGA	17 11/1	(1100)6	170	55	11115-15-15-1
XBLG-089	F:	TGTCTTGTGATCGAAATCAGG	FAM	$(AG)_7$	177	55	KX545455
11220 007	R:	ACTTCAGAGGCATTCCAGCA		(110)/	1,,,	00	
XBLG-092	F:	CTCAAGCCGATTGATGATCC	TAM	$(AG)_8$	144	55	KX545456
	R:	TCATAACATGTCACATTCAAAGGA		()0			
XBLG-097	F:	TTTGGGAAAGTCCCATGAAA	TAM	(ATC) ₆	193	55	KX545457
	R:	GGGTACAAGTGGATACAATGAGG					
XBLG-099	F:	TGCAAGGGTACATGCCATAG	ROX	$(AC)_7$	165	55	KX545458
	R:	CCAAACATTTGCCCACTTCT					
XBLG-111	F:	GAGAGGTACAACCACCACG	HEX	$(ACT)_6$	192	58	KX545459
	R:	GCCCGAAGTTAAGTAAATGGAT					
XBLG-119	F:	GCATGGTGTGTTTGGTCAAG	ROX	$(AAG)_5$	350	58	KX545460
	R:	TCTCAACAGACCCTCGTCG					

Note: T_a = annealing temperature.

polyacrylamide gel electrophoresis. We hand-selected 120 loci based on desired criteria (representative loci with different repeat unit lengths), of which 25 (20.83%) were successfully amplified and found to be polymorphic in the nine wild L. glauca populations (Appendix 1, Table 1), while 71 (59.17%) primer pairs produced no product, 21 (17.50%) amplified monomorphic markers or identical heterozygotic genotypes, and three (2.50%) produced larger or smaller products than the expected size. Forward primers of the 25 primer pairs were further labeled with fluorescently labeled nucleotides (M13: 5'-TGTAAAAC-GACGGCCAGT-3'). PCR reactions were performed in a total reaction volume of 15 µL, which contained 7.5 µL of 2× Taq PCR MasterMix (Aidlab, Beijing, China), 1.0 µL of 30 ng/µL DNA, 5.5 µL of ddH₂O, 0.5 µL of 10 µM reverse primer, 0.2 μL of 10 μM forward primer, and 0.3 μL of 10 μM fluorescent dyes (M13F-FAM, M13F-HEX, M13F-TAM, and M13F-ROX). Thermocycling program conditions included a 5-min melting step of 94°C; then 35 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 35 s; and a final extension step of 72°C for 10 min. Ten microliters each of all M13F-labeled PCR products were sent to the

Ruibo Biotechnology Center DNA Sequencing Facility (Beijing, China) for fragment analysis using an ABI 3730XL DNA Analyzer with a GeneScan 500 LIZ Size Standard (Applied Biosystems, Changping, Beijing, China). Allele genotyping was performed using GeneMarker version 2.2.0 software (Soft-Genetics, State College, Pennsylvania, USA).

Detection of SSR polymorphism and data analysis—The 25 novel polymorphic SSRs yielded 81 total alleles confirmed and genotyped via capillary gel electrophoresis. Using GENEPOP 3.2 software (Rousset, 2008) for each population, the resulting genotypic data from the capillary gel electrophoresis were analyzed to obtain standard descriptive statistics and to test for deviations from Hardy–Weinberg equilibrium (HWE) assumptions (Table 2). The total number of alleles ranged from one to four with a mean of 3.240. The observed and expected heterozygosity ranged from 0.00 to 0.90 and from 0.00 to 0.79 with averages of 0.201 and 0.479, respectively. HWE and linkage disequilibrium using Bonferroni correction were tested for every locus. Less than half of the loci (five,

I	ATN	1 (n = 10)		JGS(n = 1t)	()	Г	DZ (n = 10)	((SIG (n = 1)	()	Z	TB $(n = 10)$	()	Y	TH(n = 10)		DB	S(n = 10)		HMF	(n = 10)		TMS (n	= 10)	l
Locus	A H_{0}	H _e HWI	E^{b} A	$H_{\rm o}$ $H_{\rm e}$	$\mathrm{HWE}^{\mathrm{b}}$	Α	$H_{\rm o}$ $H_{\rm e}$	HWE ^b	Α	$H_{\rm o}$ $H_{\rm e}$	$\mathrm{HWE}^{\mathrm{b}}$	A 1	H_{\circ} H_{e}]	НWEb	Α,	H _o H _e F	łWE ^b	$A = H_{i}$, H _e H	νE	A H_{0}	H _e HN	νE	$A H_0$	H _e HWE	l ස l
XBLG-013	3 0.3() 0.54 **	1	0.00 0.00	Μ	2 C	0.00 0.19	***	4	0.00 0.69	***	4 0.	00 0.69	* *	4	20 0.67	* *	2 0.0	0 0.51 *	*	2 0.00	0.34 *	*	4 0.00 0	.61 ***	
XBLG-033	1 0.00	0.00 M	3	0.00 0.65	* * *	2	0.40 0.51	n.s.	3	0.00 0.59	***	1 0.	.00 0.00	Μ	1 0.	00 0.00	Μ	2 0.1	0 0.39	*	2 0.00	0.51 *	*	2 0.20 0	.19 n.s.	
XBLG-036	2 0.3() 0.52 n.s.		0.20 0.19	n.s.	2	0.00 0.19	***	3	0.00 0.59	***	2 0.	.10 0.27	*	2	.00 0.19	***	2 0.0	0 0.19 *	*	2 0.10	0.10 n	l.S.	3 0.10 0	.68 ***	,,,
XBLG-049	3 0.2() 0.56 n.s.		0.40 0.69	n.s.	3 (0.10 0.65	***	3 (0.10 0.53	* *	2 0.	.20 0.19	n.s.	2	.10 0.27	*	3 0.5	0 0.68 n	s.	3 0.10	0.64	*	2 0.00 0	.51 ***	
XBLG-051	3 0.2() 0.61 ***	5	0.70 0.48	n.s.	3 (0.90 0.65	*	2	0.30 0.27	n.s.	3 0.	.30 0.43	n.s.	3 0.	.30 0.54	n.s.	3 0.0	0 0.36 *	*	2 0.20	0.34 n	l.S.	3 0.10 0	.68 n.s.	
XBLG-053	3 0.4() 0.47 n.s.		0.40 0.34	n.s.	3 (0.60 0.69	n.s.	5	0.00 0.51	***	2 0.	.00 0.44	***	3 0.	.10 0.62	***	2 0.2	0 0.19 n	s.	3 0.10	0.59 *	*	3 0.50 0	.43 n.s.	
XBLG-055	3 0.3(* 69.0 (3	0.60 0.57	**	2	0.00 0.19	* **	2	0.00 0.51	* *	2 0.	.00 0.19	*	3 0.	.00 0.48	***	3 0.3	0 0.54	*	3 0.00	0.65 *	*	3 0.80 0	* 69.	
XBLG-056	1 0.00	0.00 M	-	0.00 0.00	Μ	2	0.00 0.19	* **	3 (0.20 0.35	n.s.	2 0.	.00 0.19	***	2	.30 0.48	n.s.	2 0.0	0 0.19 *	*	2 0.00	0.34 *	*	2 0.00 0	.34 ***	
XBLG-058	3 0.3() 0.53 ***	4	0.60 0.50	n.s.	2	0.70 0.48	n.s.	4	0.66 0.66	n.s.	4 0.	.30 0.72	***	4	.30 0.62	***	4 0.4	, 69.0 0	*	3 0.20	0.61	*	4 0.20 0	.65 **	
XBLG-060	3 0.3() 0.43 ***		0.00 0.00	Μ	4	0.20 0.36	*	4	0.70 0.79	n.s.	3 0.	.20 0.57	***	4	.20 0.36	*	2 0.2	0 0.34 n	s.	2 0.10	0.10 n	l.S.	2 0.10 0	.27 *	
XBLG-062	1 0.00	0.00 M	-	0.00 0.00	Μ	1	0.00 0.00	Σ	3	0.30 0.28	n.s.	2 0.	.10 0.10	n.s.	4	.30 0.44	*.	1 0.0	0 0.00	V	1 0.00	0.00	M	3 0.20 0	.19 n.s.	
XBLG-063	2 0.2() 0.44 n.s.		0.10 0.43	* * *	2	0.00 0.19	***	5	0.20 0.21	*	3 0.	.30 0.59	***	3 0.	.30 0.56	n.s.	3 0.2	0 0.35 n	s.	2 0.00	0.51 *	*	2 0.10 0	.52 **	
XBLG-066	1 0.00	0.00 M	-	0.00 0.00	Μ	2	0.00 0.19	* * *	5	0.10 0.10	n.s.	1 0.	.00 0.00	Μ	2	.10 0.10	n.s.	2 0.9	0 0.52	*	2 0.50	0.39 n	l.S.	2 0.50 0	.39 n.s.	
XBLG-073	1 0.00	0.00 M	2	0.20 0.19	n.s.	2	0.00 0.19	***	1	00.0 00.0	Μ	1 0.	.00 0.00	Μ	2	.10 0.27	n.s.	1 0.0	0 0.00	V	2 0.30	0.27 n	l.S.	2 0.10 0	.10 n.s.	
XBLG-076	1 0.00	M 00.0 (-	0.00 0.00	Μ	2	0.60 0.44	n.s.	5	0.50 0.39	n.s.	2 0.	.30 0.27	n.s.	2	40 0.34	n.s.	3 0.4	0 0.35 n	s.	2 0.50	0.39 n	l.S.	3 0.20 0	.54 n.s.	
XBLG-082	3 0.2() 0.19 n.s.		0.20 0.19	n.s.	3 (0.30 0.42	n.s.	3 (0.50 0.54	n.s.	3 0.	.30 0.62	*	3 0.	.30 0.59	***	2 0.0	0 0.34 *	*	3 0.30	0.62	*	3 0.20 0	.** 99.	
XBLG-083	3 0.3() 0.54 n.s.		0.30 0.27	n.s.	3 (0.00 0.36	***	3	0.30 0.58	*	3 0.	.10 0.42	*	3 0.	.10 0.56	*	2 0.0	0 0.19 *	*	2 0.10	0.10 n	l.S.	2 0.00 0	.19 ***	,,,
XBLG-084	2 0.1() 0.52 *	2	0.00 0.44	***	2	0.00 0.53	***	3	0.00 0.61	***	2 0.	40 0.51	n.s.	2	.30 0.48	n.s.	2 0.0	0 0.34 *	*	3 0.10	0.53 *	*	3 0.20 0	.54 n.s.	
XBLG-086	1 0.00	M 00.0 (1	0.00 0.00	Μ	2	0.20 0.34	n.s.	5	0.10 0.27	*	2 0.	.20 0.34	n.s.	2	.00 0.51	***	3 0.2	0 0.54	*	3 0.20	0.65	*	2 0.30 0	.27 n.s.	
XBLG-089	2 0.3() 0.27 n.s.		0.00 0.00	Μ	1 (0.00 0.00	Σ	1	00.0 0.00	Μ	2 0.	.40 0.51	n.s.	1 0.	00.0 00.	Μ	1 0.0	0 0.00	V	2 0.60	0.44 n	l.S.	2 0.20 0	.53 *	
XBLG-092	2 0.4() 0.51 n.s.		0.80 0.57	n.s.	2 (0.00 0.19	***	4	0.40 0.76	*	2 0.	.10 0.39	*	3 0.	50 0.67	*	2 0.2	0 0.34 n	s.	3 0.10	0.51	*	4 0.50 0	* 99.	
XBLG-097	3 0.6() 0.68 *	3	0.90 0.59	n.s.	3 (0.10 0.28	***	3	0.40 0.48	***	3 0.	.20 0.57	n.s.	4	30 0.60	n.s.	3 0.2	0 0.19 n	s.	3 0.20	0.19 n	l.S.	4 0.70 0	.66 n.s.	
XBLG-099	3 0.2() 0.57 ***	3	0.20 0.56	n.s.	2 (0.10 0.10	n.s.	5	0.00 0.51	***	3 0.	.00 0.65	***	3 0.	.10 0.69	***	2 0.1	0 0.39	*	3 0.50	0.47 n	l.S.	4 0.30 0	.75 ***	,,,
XBLG-111	3 0.2() 0.36 ***	3	0.10 0.43	* * *	3 (0.60 0.65	*	3	0.20 0.62	*	2 0.	.00 0.19	***	2	.20 0.53	*	2 0.0	0 0.44 *	*	3 0.20	0.28	*	2 0.30 0	.52 n.s.	
XBLG-119	2 0.3() 0.48 n.s.		0.10 0.39	*	2	0.10 0.48	*	5	0.30 0.39	n.s.	2 0.	.10 0.52	*	2	.20 0.53	*	2 0.1	0 0.52	*	2 0.10	0.52	*	2 0.00 0	.19 ***	,,,
Mean 2	.20 0.2(0.36	2.0	0 0.23 0.30		2.14 (0.20 0.34		2.64 (0.21 0.46		2.32 0.	.14 0.37	. 4	2.64 0	.22 0.44	. 1	2.24 0.1	6 0.34	0	.40 0.18	0.40	0	72 0.27 0	.47	
Note: $A =$	= total 1	number of	allele	$3S; H_{a} = exp$	vected h	etero	zygosity	$H_0 =$	obser	ved hete	rozygo	sity; F	HWE = H	Hardv-	Weint	berg equil	ibriun	n; n = r	number of	indiv	/iduals s	ampled				1
^a Locality	and ve	oucher infe	ormat	tion are pro	vided ir	ιApp	bendix 1.	>			,)	•		,		•						•				
^b Asterisk	s indic	ate signific	cant d	leviation fre	om Har	dv-Vb	Veinberg	equilit	ninm	(*P < 0)	05, **!	⁹ < 0.(01, ***P	>< 0.00	11); N	1 = mono.	morph	ic; n.s.	= not sig	nifica	int.					
)				,)	•		,							•		,							

TABLE 2. Descriptive statistics of the 25 newly developed polymorphic microsatellites of *Lindera glauca.*^a

http://www.bioone.org/loi/apps

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TABLE 3.	Cross-amplification results for	he 25 polymorphic cDN	JA-SSR loci developed for	r <i>Lindera glauca</i> in 1	0 individuals of L. communis. ^a
	1	1 2 1	1	0	

Locus	LC001	LC002	LC004	LC005	LC009	LC010	LC011	LC019	LC021	LC022
XBLG-013	0	0	1	0	0	0	1	0	1	0
XBLG-033	0	1	1	0	1	0	0	0	0	0
XBLG-036	1	0	0	1	0	1	1	0	0	0
XBLG-049	1	1	1	1	0	1	1	1	0	1
XBLG-051	1	1	1	1	1	1	1	1	1	1
XBLG-053	1	1	1	1	1	1	1	1	1	1
XBLG-055	1	1	1	1	1	1	1	0	1	1
XBLG-056	1	1	1	1	0	1	1	1	1	1
XBLG-058	0	0	1	1	1	1	1	1	1	1
XBLG-060	1	1	1	1	0	1	1	1	1	1
XBLG-062	1	1	1	1	1	1	1	1	1	1
XBLG-063	1	1	1	1	1	1	1	1	1	1
XBLG-066	1	1	1	1	1	1	0	1	1	0
XBLG-073	1	1	1	0	1	1	1	1	0	1
XBLG-076	1	1	1	1	1	1	0	1	1	1
XBLG-082	1	0	1	1	1	1	1	1	1	1
XBLG-083	1	1	1	0	0	1	0	1	1	1
XBLG-084	1	0	1	1	1	0	1	1	1	0
XBLG-086	1	1	1	1	0	1	1	0	1	1
XBLG-089	1	1	1	1	0	1	1	1	1	1
XBLG-092	0	1	1	1	1	0	1	1	1	1
XBLG-097	1	0	1	0	1	1	1	1	1	1
XBLG-099	0	1	1	1	1	1	1	1	1	1
XBLG-111	1	0	1	0	1	1	1	0	1	0
XBLG-119	0	0	1	0	0	0	1	0	0	0

Note: 1 = successful amplification; 0 = failed amplification.

^aLC = population names of *Lindera communis*. Samples were collected in Longjiang County, Yunnan Province, China (geographic coordinates: 24°46′33″N, 98°39′25″E; elevation: 1219 m) and identification codes are kept at the Laboratory of Systematic Evolution and Biogeography of Woody Plants, School of Nature Conservation, Beijing Forestry University, Beijing, China.

five, 12, nine, eight, seven, eight, seven, and seven loci in populations ATM, JGS, LDZ, SJG, NTB, YTH, DBS, HMF, and TMS, respectively) showed significant departure from HWE (P < 0.001). Significant linkage disequilibrium was not detected between any pair of loci (P < 0.001).

Cross-species amplifications—The 25 primers were tested in 10 individuals of *L. communis* Hemsl. under the same PCR conditions as above. All 25 were found to amplify in at least 21 of the species (Table 3).

CONCLUSIONS

In the current study, we developed 25 novel cDNA-SSR markers that were highly polymorphic in *L. glauca* and used these markers to successfully investigate genetic distances within nine wild populations of *L. glauca*. The collection of SSRs presented herein provide a means to assess genetic diversity and to further investigate large-scale and fine-scale population genetic structure in *L. glauca*. These markers may also be useful for germplasm identification and breeding programs in both this species and other species in the genus *Lindera* Thunb.

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		Geographic	coordinates			·
Population	Sample accession no.	Latitude	Longitude	Elevation (m)	Province in China	County
ATM	A14-10	31°13′30″N	115°51′35″E	646-834	Anhui	Jinzhai
JGS	J13-09	31°52′15″N	114°05'13"E	203-317	Henan	Xinyang
LDZ	L14-04	31°56′47″N	114°15′26″E	154-261	Henan	Dongzhai
SJG	S14-10	31°44′58″N	115°32'29"E	243-476	Henan	Shangcheng
NTB	N14-04	32°19′45″N	113°25′24″E	241-256	Henan	Tongbai
YTH	Y14-04	31°03′24″N	115°51′54″E	647-734	Hubei	Yingshan
DBS	D14-09	31°00'32"N	115°50'12"E	834-1003	Hubei	Yingshan
HMF	H14-09	28°26′51″N	113°00'22"E	224-257	Hunan	Wangcheng
TMS	T14-09	30°19′28″N	119°26'56"E	359-432	Zhejiang	Linan

APPENDIX 1. Location and sampling information for Lindera glauca individuals used in this study.^a

Note: *n* = number of individuals sampled.

^aSample accession numbers refer to voucher specimens deposited in the Herbarium of the Beijing Forestry University (BJFC), Beijing, China; geographic coordinates and elevation were obtained with a portable GPS receiver.